

The Classification of Aflatoxin Contamination Level in Cocoa Beans using Fluorescence Imaging and Deep learning

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Abstract—Aflatoxin contamination in cacao is a significant problem in terms of trade losses and health effects. This calls for the need for a non-invasive, precise, and effective detection strategy. This research contribution is to determine the best deep-learning model to classify the aflatoxin contamination level in cocoa beans based on fluorescence images and deep learning to improve performance in the classification. The process involved inoculating and incubating *Aspergillus flavus* (6mL/100g) to obtain aflatoxin-contaminated cocoa beans for 7 days during the incubation period. Liquid Mass Chromatography (LCMS) was used to quantify the aflatoxin in order to categorize the images into different levels including “free of aflatoxin”, “contaminated below the limit”, and “contaminated above the limit”. 300 images were acquired through a mini studio equipped with UV lamps. The aflatoxin level was classified using several pre-trained CNN approaches which has high accuracy such as GoogLeNet, SqueezeNet, AlexNet, and ResNet50. The sensitivity analysis showed that the highest classification accuracy was found in the GoogLeNet model with optimizer: Adam and learning rate: 0.0001 by 96.42%. The model was tested using a testing dataset and obtain accuracy of 96% based on the confusion matrix. The findings indicate that combining CNN with fluorescence images improved the ability to classify the amount of aflatoxin contamination in cacao beans. This method has the potential to be more accurate and economical than the current approach, which could be adapted to reduce aflatoxin's negative effects on food safety and cacao trade losses.

Keywords—Aflatoxin; Cocoa Beans; Deep Learning; Fluorescence Images.

I. INTRODUCTION

Aflatoxin exposure has a significant detrimental impact on consumer health and the global economy. A recent review by [1] estimated that 60% - 80% of the world's food crops were contaminated by aflatoxin. Aflatoxin can harm human health, leading to death [2-3], cancer [4-5], immune system disruption [6], and stunted child growth [7-8]. *Aspergillus flavus* produces aflatoxin, a secondary metabolite that thrives in uncontrolled processing and storage condition [9]. The contamination needs to be controlled early due to its thermostable nature, which can complicate the removal process from the supply chain [10]. This is necessary because of the recent increase in the market awareness of product

quality [11] as well as the existence of strict regulations governing the maximum intake of aflatoxin by international institutions such as the 20 ppb limit imposed by the European Commission [12].

One of commodity that can become contaminated with aflatoxin is cocoa [13]. Cocoa beans are one of the important commodities in the world as indicated by their retail value reaching USD 100 billion in 2021. The total trade volume of cocoa and its products in the year 2018-2019 was also reported to be \$50.9 billion with a 2.12% increase in export value [14]. However, several challenges have been identified in relation to the export of cocoa such as the reduction in the quality due to the risk of mycotoxins [15].

The detection of aflatoxin was observed to currently rely on the application of chemical approaches involving high costs, destructive processes, sophisticated equipment, and labor expertise [16] such as ELISA [17], HPLC [18], and LCMS [19]. Another method is computer vision which involves utilizing different image acquisition technologies such as Color imaging [20], Near-infrared (NIR) Spectroscopy [21-22], Mid-infrared (MIR) Spectroscopy [23-24], Near-infrared (NIR) Hyperspectral Imaging [25-26], and even X-Ray Imaging [27] to detect mycotoxin. However, these image acquisition technologies have some limitations. For instance, color imaging has a limited electromagnetic range, making it unable to detect mycotoxin contamination at an early stage, despite its high detection accuracy of up to 89% [28]. Hyperspectral imaging has high accuracy [29] but requires expensive equipment [30]. This requires an image acquisition approach that is simple but has high accuracy.

Ultraviolet (UV) induced fluorescence imaging is an alternative technique involving the utilization of the excitation-emission properties of fluorescence light. This is possible because every organic material, including aflatoxin, exhibits different fluorescence properties [31]. Previous research already showed that aflatoxin has an excitation wavelength of 365 nm and an emission wavelength of 455 nm [32]. The fluorescence properties were used in the hyperspectral fluorescence imaging approach to detect the aflatoxin in maize [33]. However, this approach requires relatively expensive equipment, necessitating a relatively



simpler approach by Rotich [34], 365 nm LEDs were used as the excitation light source in a color camera to capture the optical characteristics of fluorescence images produced by UV light in the visible region. The target fluorophores were observed to have been able to emit in the visible region thereby, enabling the use of simplified imaging techniques. This indicates that there is potential for using fluorescence imaging in this study.

Convolutional Neural Network (CNN) is an approach commonly used for image classification [35-37]. This was further confirmed by the findings of recent research that machine learning is increasingly being applied in the field of agriculture, particularly to identify the quality of agricultural products. The phenomenon was associated with the machine learning's capabilities in the classification process, which is a recent development in computer vision [38]. Hendrawan [39] classified Indonesian coffee types using a pre-trained network with a testing accuracy of 99.6%. Momeny [40] also employed the Inception-v4 CNN to grade and detect counterfeit saffron with an accuracy of 99.5%. CNNs can not only be used for classifying reflectance images but are also well-suited for handling fluorescence images. The findings of the research support this statement. For example the classification performance of fluorescence images was reported to be better than that of reflectance images, as shown by Wei [41] where fluorescence images had a higher accuracy of 97.5% in the process of classifying tea leaves.

This research contribution is to determine the best deep-learning model to classify the aflatoxin contamination level in cocoa beans based on fluorescence images and deep learning to improve performance in the classification. The results are expected to offer an alternative, accurate method for classifying aflatoxin contamination levels in cocoa beans, which possible adapted to reduce aflatoxin's negative effects on food safety and cacao trade losses.

II. MATERIALS AND METHOD

A. Materials

The materials used included pure strain *A. flavus* (Inacc F44) was collected from Lembaga Ilmu Pengetahuan Indonesia (LIPI) microbiology laboratory, Cibinong, Indonesia. Forastero cocoa beans that had been fermented for 5 days with a shelf life of 3 months in a warehouse from the Pusat Penelitian Kopi dan Kakao, Jember, Indonesia. The fermented beans were unbroken, free of mycotoxin contamination and had a low moisture content of 6%. The chemicals used were aqua dest, NaCl, methanol, alcohol, and potato dextrose broth (PDB) media.

The hardware used was image acquisition mini studio consisting of a Canon EOS 700D DSLR camera with dimensions of 133×100×79 mm, equipped with a PL filter. The camera had 18MP resolution specifications with a CMOS sensor, ISO 100-12800, a shutter speed of 30-1/4000 seconds, and Full HD video. 22.3×14.9 mm CMOS sensor size. UV Lamp Model: LDR2-100UV3-365-N, input 24V(DC)/23W, manufactured by CCS Inc as a UV light source. The power supply had 30V/5A Specification with code MDB-K305D. A LED Pulse Controller, Brand GARDASOFT with model PP820C, which was equipped with 8 channels that can

regulate a maximum current of 20A was used, with an input of 12-48V. A mini studio frame was assembled from several angles with a thickness of 2 mm combined using 10mm nuts and a Lenovo laptop with Intel Core i5-7200u specifications, 2.50GHz CPU 2.71GHz, and 16GB memory. Aflatoxin quantification using Liquid Chromatography Mass Spectrometry (LCMS)/MS brand (Hitachi L 6200).

B. Method

The research's outline stages can be seen in Fig.1. As seen in Fig. 1, this study uses the inoculation and incubation of *A. flavus* to produce aflatoxins-contaminated cocoa beans. The fluorescence image acquisition process is carried out and the last step is the implementation of a pre-trained CNN model. The following provides a detailed explanation of each of the aforementioned stages.

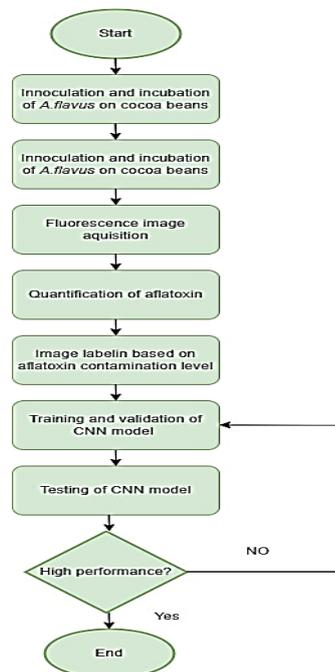


Fig. 1. Research outline

1) Fungus Inoculum

Fungal inoculation is carried out to obtain cocoa beans contaminated with aflatoxin. Fungal inoculum was obtained by culturing pure *A. flavus* in 10 mL of nutrient broth (DIFCO 234000) and incubating it at room temperature for 3 days. In 100 mL of NB media, the culture was revived, re-inoculated, and incubated at room temperature for three more days. To create a stock inoculum for the vaccination of cocoa beans, the resulting culture was then incubated for an additional three days at room temperature in 1000 mL of NB media. 60 mL/kg of cocoa beans were added to the fungal inoculum, which was then incubated with *A. flavus* at 30° C and 90% RH until day 7.

2) Measurement of Aflatoxin Levels

The concentration of aflatoxin in the samples was measured using LCMS. The LC system consisted of: (i) a Shimadzu pump model LC-10AD (Shimadzu, Kyoto, Japan); (ii) a fluorescence detector (RF-10AXL) (Shimadzu); (iii) an auto-injector (SIL-10A, Shimadzu); and (iv) a control

system (SCL-10A, Shimadzu), as well as a mass spectrometry detector with an orthogonal ESI nozzle (model Agilent 1100MSD SL). The Aflatoxin (AF) standard and pure samples were dissolved in 1 ml of mobile phase solution, which was then added to the LCMS/ESI in 6 μ L. All of the tested AF contained protons, $[M+H]^+$, with a residence time of 1,000 ms per ion. The test tube containing the pure sample or the aflatoxin working standard was supplemented with 0.1 mL of trifluoroacetic acid (TFA) for LC analysis. The selected ions (m/z) monitored for the target were 313 for AFB₁, 315 for AFB₂, 329 for AFG₁, and 331 for AFG₂ [42]. The tube was vortexed, diluted with 0.9 ml of an acetonitrile-water solution (1:9), and allowed to sit in the dark for 15 minutes at room temperature. The resulting solution was then tested for reverse-phase LC analysis using 20 μ L. Using electron ionization in multiple reaction monitoring modes, aflatoxin was discovered. The results of the LC/MS-MS data analysis gave the extract's compounds' molecular weight and chromatogram in the form of peaks, allowing for the calculation of how many compounds were total in each sample. For each measurement, three times were performed.

3) Image Acquisition

Canon EOS 700D camera was used to capture images of cocoa beans for up to seven days following the inoculation of *A. flavus* to obtain images of cocoa beans in 3 different aflatoxin contamination classes (aflatoxin free, below threshold, and above threshold). 400 \times 600 pixel JPG files were used to save the generated images, then the image is resized to 224 \times 224 pixels as the input image to the CNN [43]. The image was captured in a studio that featured a 365 nm UV LED lamp made by CCS Inc. in Japan. The UV lamp was positioned 350 mm away from the sample unit, emitting 6.9 Wm² of radiation on average. Additionally, a UV bandpass filter was placed in front of the UV light source and the camera to block out the light that was reflected. It performs similarly to a UV cut filter but only lets 0.3% of 365 nm to pass through. In the subunit of image acquisition, a high-resolution CMOS camera of 5 \times 3078 pixels was used, namely the EOS Kiss \times 7 (Canon Inc., Japan) with ISO 200, F-5.6, and manual exposure of 1/3 second for fluorescence image recording placed 450 mm from the sample location. The image acquisition unit measures 18.5 cm \times 18.5 cm \times 29 cm and is connected to the light source through an optic fiber, as shown in Fig. 2.

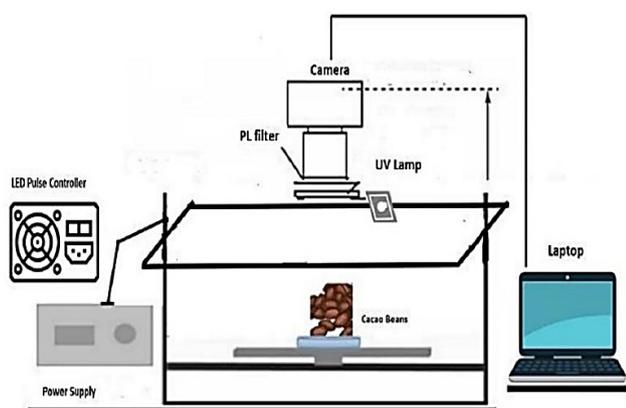


Fig. 2. Image acquisition unit

Each cocoa bean is spaced approximately 5 cm apart in the arrangement. After seven days of image acquisition, 300 fluorescence images with 12 cocoa beans each were obtained. Based on the findings of the LCMS test, the image is then classified into three classes of contamination.

4) Deep Learning Modeling

The model was created using deep learning to classify the aflatoxin contamination level in cocoa beans. Four different pre-trained CNN types were used in the process: SqueezeNet [44], GoogLeNet [45], ResNet50 [46], and AlexNet [47]. The CNN architecture for the classification is presented in the following Fig. 3.

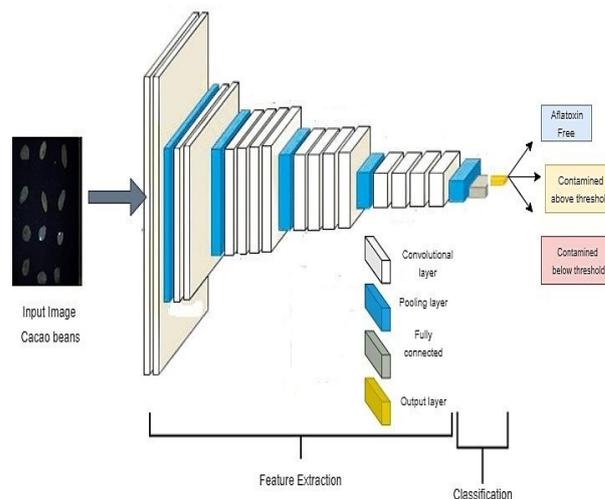


Fig. 3. CNN structure for aflatoxin contamination level classification

As seen in Fig. 3, the cocoa image becomes input to the CNN model and then convolution and pooling are used to extract features. In the fully connected layer, classification is carried out so that the output is three classes of aflatoxin contamination in cocoa beans. Each pre-trained CNN was applied using several parameter settings such as optimizer (Adam, SGDm, RMSProp) [48], initial learning rate = 0.0001 and 0.00005 [49], epoch 20 of minibatch size 20 [50], L2Regularization = 0.00001 [51], momentum = 0 [52], learning rate drop period = 10 [53], and learner drop factor = 0 [54]. The value of sequence padding is 0 [55]. Padding affects network functionality and has a significant impact on performance and accuracy [56]. After validation, the CNN models were tested using 20 new images for each aflatoxin contamination level category. Moreover, the confusion matrix method [36] was used to determine the classification accuracy of the testing dataset in order to evaluate the performance of the CNN models. The architecture of the pre-trained CNN used is presented in the following Fig. 4.

The ability of pre-trained CNNs to expedite and streamline the training process is one of its key advantages. The pre-trained model's weights and features can be used as a starting point and adjusted to particular task, rather than having to be started from scratch [57]. Each pre-trained network possess various architectures, parameter counts, and levels of complexity, all of which affect the accuracy and classification rate [58].

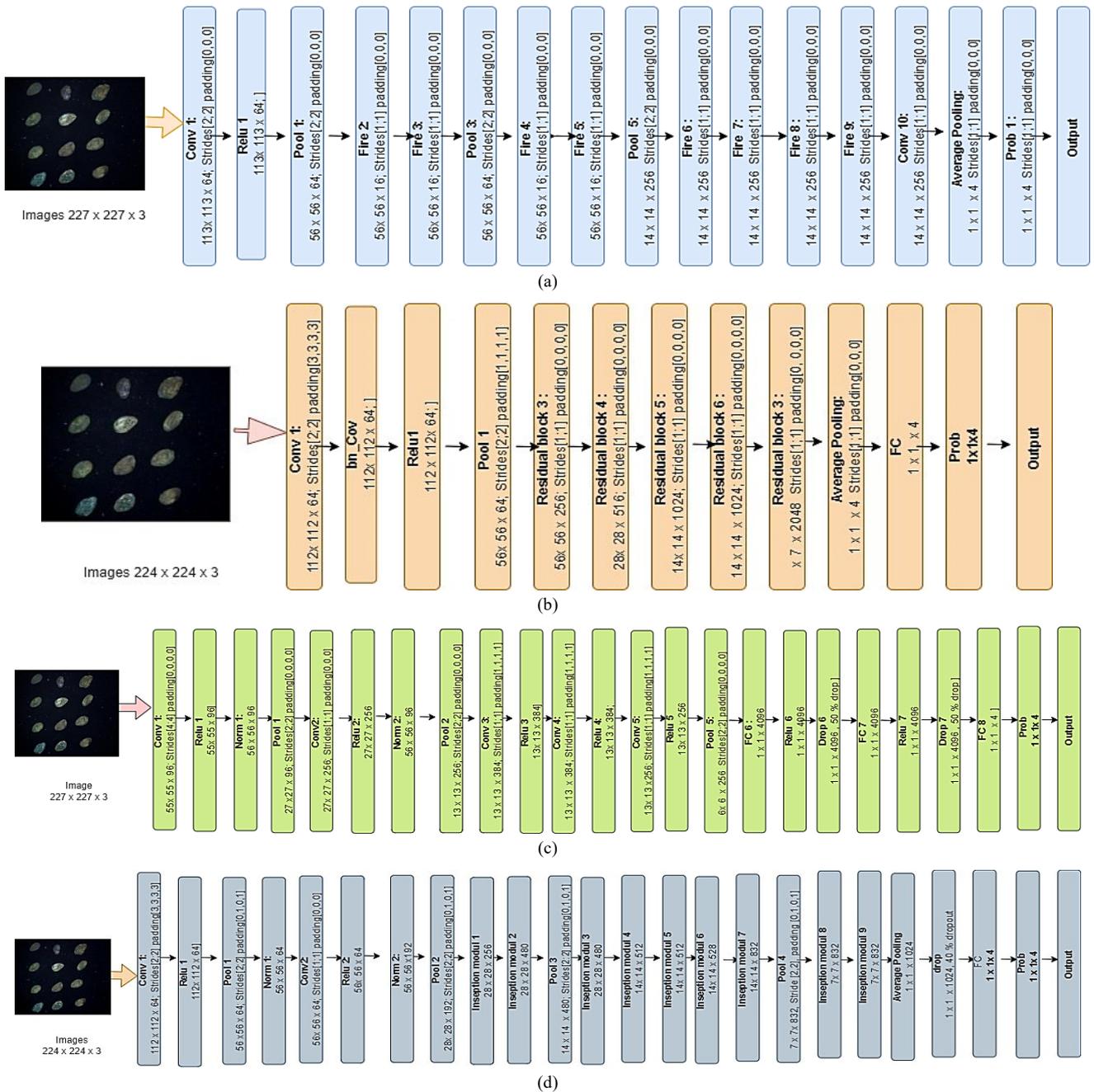


Fig. 4. Schematic representation of the CNN models (a) SqueezeNet, (b) ResNet50, (c) AlexNet, (d) GoogleLeNet

III. RESULTS AND DISCUSSION

A. Quantification of Aflatoxin Level in Cocoa Beans Inoculated with *Aspergillus Flavus*

In this study analyzed total aflatoxin (AF) which is the total of aflatoxin B1 (AFB₁), aflatoxin B2 (AFB₂), aflatoxin G1 (AFG₁) and aflatoxin (AFG₂). The detection limit of the LC-MS/MS test in this research was 1 ppb (part per billion). Samples were considered positively contaminated when the aflatoxin concentration was \geq Limit of detection (LOD). Fig. 5 indicated that the control did not detect aflatoxin at the limit of Reporting limit (RL) = 1 μ g/kg, while the Aflatoxin (AF) inoculation treatment on days 1-7 did not detect AFB₂ and AFG₂. Furthermore, AFB₁ and AFG₁ levels increased during

the observation period, with AFG₁ showing the largest increase in aflatoxin level at 83.10 ppb on day 7, followed by AFB₁ at 9.77 ppb on day 7. Aflatoxin level was significantly different in each observation. The higher concentration of AFG₁ than AFB₁ in this research was in line with [16], where aflatoxins in cocoa beans were analyzed in several regions in Brazil. The results showed an incidence of aflatoxin level in cocoa beans aflatoxin G1 higher than B1. However, some findings revealed that AFB₁ and AFG₁ levels were equal or AFB₁ level was higher than AFG₁. Compared to the incidence of aflatoxin contamination in other commodities such as peanuts, various reports obtained a higher level of AFG₁ than AFB₁[58], while others detected elevated AFB₁ than AFG₁[59][60]. Comparable concentration patterns of

AFB₁ and AFG₁ were also reported in beans from Brazil [61] and Burkina Faso [62].

The aflatoxin concentration in cocoa beans was found to exceed the threshold on day 3 after inoculating. Hruska [33] discovered that the aflatoxin level in corn kernels inoculated with *A. flavus* fungus exceeded the threshold on day 7. This condition occurred due to various factors that affected mold growth and aflatoxin production. Intrinsic factors included water activity [59], pH [60], redox potential [61], substrate [62], inhibitors [63], and osmotic pressure [64]. Meanwhile, extrinsic factors were related to environmental conditions such as temperature and humidity [65]. The incubator's temperature and relative humidity (RH), the amount of media added, and cocoa bean nutrient content (substrate) were all factors under control in this study that affected aflatoxin levels. In order to drive the growth of mold and the generation of aflatoxin, the incubator's temperature and RH were set to 30 °C and 90 RH, respectively. Biotrop [66] reported that

compared to temperatures of 40° C and 80 RH, a temperature of 30 °C with 90 RH was better for the development of molds and the production of aflatoxin. This is due to the mold's inability to grow at 40° C with a 70% RH.

Nutrients from the substrate such as lipids, carbon, nitrogen, and amino acids are among the factors that can affect aflatoxin production [67]. Cocoa beans, which are composed of cocoa butter (50%), mainly stearic acid, oleic acid and palmitic acid, containing protein (11%), including glutamate, arginine and leucine provide an ideal commodity type for mold and aflatoxin development. Glutamate is a source of arginine and proline [68], which are nitrogen sources to stimulate aflatoxin production [9]. The highest aflatoxins are induced by glutamate and aspartate, followed by arginine [62]. Nitrogen in the form of nitrite and nitrate also increases the level of aflatoxin production [69]. Moreover, aflatoxin biosynthesis is also triggered by lipophilic epoxy fatty acids and induced by ergo-sterol oxidation [70].

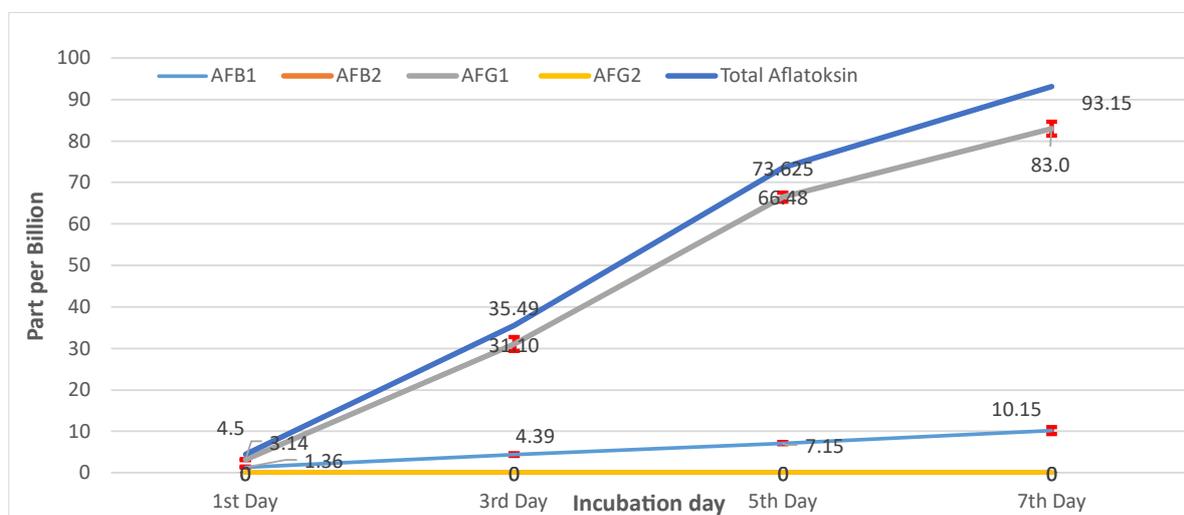


Fig. 5. Changes in aflatoxin levels after inoculation of *A. flavus* on cocoa beans

B. Image acquisition and analysis

Fluorescence images were acquired for this study by excitation of UV light at 365 nm wavelength. Airborne mycotoxin is detected using an excitation source of 360–370 nm; and 435–440 nm for substrate in food materials [71]. Furthermore, the fluorescence image of cocoa beans obtained is shown in Fig. 6 and Fig. 7.

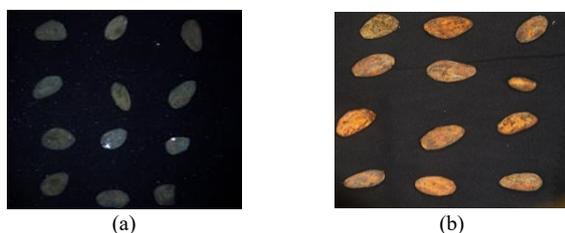


Fig. 6. Difference of reflectance and fluorescence imaging (a) Reflectance image (b) Fluorescence image

Fig. 6 shows how the reflectance and fluorescence images of aflatoxin-contaminated cocoa beans differ from one another. While there are some cocoa beans in the fluorescence image that excite blue light, as shown in Fig. 6 (b), there are

no discernible differences between the reflectance images of the coffee beans.

In Fig. 7, cocoa beans image can be observed with blue UV light emission on day 7. This emission occurs when fluorophores are excited by UV light at a specific wavelength, resulting in a longer wavelength. AFB₁ and AFB₂ have blue fluorescent colors, while AFG₁ and AFG₂ have green colors [72]. On the other hand, Fig. 6(b) shows that cocoa beans contaminated with aflatoxins are below the threshold at which, in theory, they should excite light as a sign of aflatoxin contamination; however, none of the cocoa beans exhibit light excitation. However, cocoa beans that produce excitation are not an absolute indicator of the presence of aflatoxin. It should be noted that aflatoxins are not the only ones responsible for UV light emission. According to Gao [73], UV fluorescence light may also be emitted by oil in grains and dust in the air. It's possible that accuracy will decrease if aflatoxins are only detected using blue-green fluorescence under UV light. Therefore, in the subsequent stage, deep learning approaches were employed to improve the accuracy of the detection process.

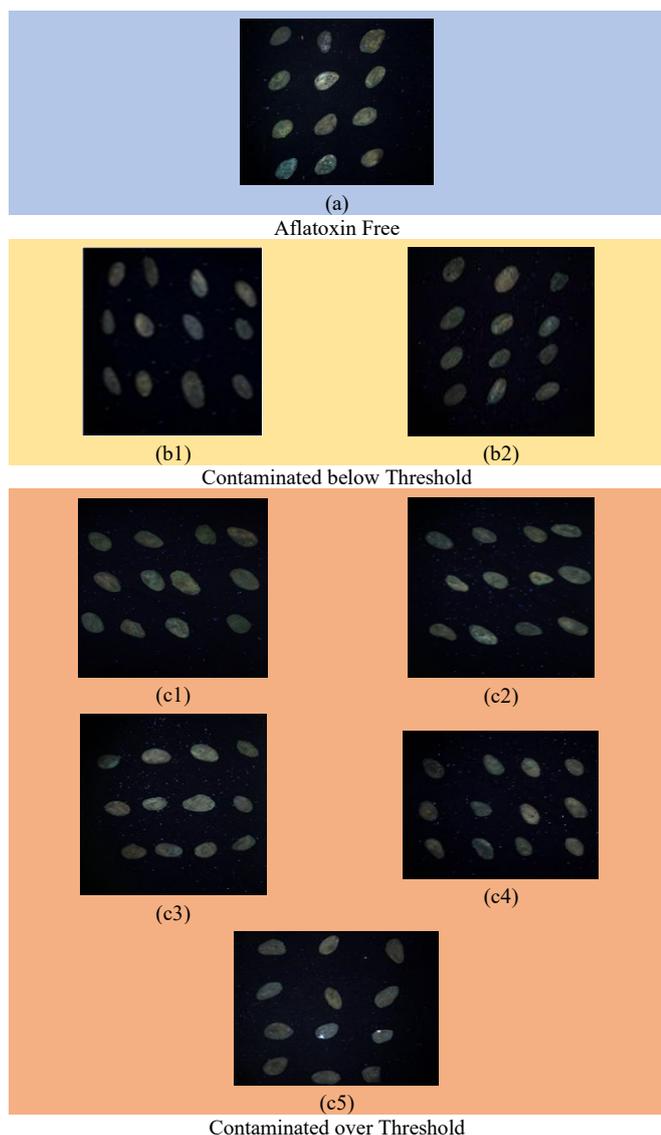


Fig. 7. Fluorescence image of cocoa beans from *A. flavus* inoculation on days 1-7, a) Control, (b1) Day 1, (b2) day 2, (C1) day 3, (c2) day 4, (c3), day 5, (c4) day 6, (c5) day 7

C. CNN Performances

This research was conducted using four pre-trained CNN models including AlexNet, GoogLeNet, ResNet50, and SqueezeNet to classify the aflatoxin contamination level in cocoa beans. Moreover, sensitivity analysis was conducted through different variations of optimization methods such as Adam, SGDm, and RMSProp as well as initial learning rates set at 0.0001 and 0.00005. The performance of these models is presented in the following Table I.

The pre-trained CNN models were observed to have produced different accuracy values with the lowest, 76.69%, recorded for GoogLeNet, optimizer: SGDm, learning rate: 0.00005 and the highest, 96.42%, for GoogLeNet, optimizer: Adam, learning rate: 0.0001. This means GoogLeNet, optimizer: Adam, learning rate: 0.0001 was the best combination to classify the level of aflatoxin contamination in cocoa beans using fluorescence images. Moreover, the average accuracy based on the type of pre-trained network showed that ResNet50 had the highest average accuracy of 91.48%, followed by AlexNet at 92.8%, GoogLeNet at

87.2%, and SqueezeNet at 87.15%. This was observed to be consistent with the findings of Liu [74] that ResNet has better accuracy than other models such as VGG-16 and MobileNet during the process of comparing the CNN models for fruit classification. However, it was discovered from Table I that ResNet50 required the longest training time of 58.2 minutes while AlexNet has the shortest at approximately 28.2 minutes.

This was likely due to ResNet50 having a higher depth of 50 layers which was more than AlexNet with 5 layers, SqueezeNet with 18 layers, and GoogLeNet with 22 layer. There are more weights and biases to update during training when the parameter size is larger. This may result in longer training times for the model by making it more complex and difficult to optimize [75].

TABLE I. COMPARISON OF THE ACCURACY OF SEVERAL PRE-TRAINED CNN ARCHITECTURES DEVELOPED TO CLASSIFY THE AFLATOXIN CONTAMINATION LEVEL USING FLUORESCENCE IMAGES

Architecture	Optimizer	Learning rate	Accuracy (%)	Time (minutes)
SqueezeNet	SGDm	0.00005	84.59	34
	Adam	0.00005	87.22	35
	RMSprop	0.00005	87.22	35
	SGDm	0.0001	85.71	35
	Adam	0.0001	93.98	35
	RMSprop	0.0001	84.21	34
AlexNet	SGDm	0.00005	83.46	23
	Adam	0.00005	90.25	22
	RMSprop	0.00005	89.47	23
	SGDm	0.0001	87.42	22
	Adam	0.0001	95.95	28
	RMSprop	0.0001	89.47	23
GoogLeNet	SGDm	0.00005	76.69	49
	Adam	0.00005	93.98	49
	RMSprop	0.00005	90.00	48
	SGDm	0.0001	81.2	48
	Adam	0.0001	96.42	51
	RMSprop	0.0001	90.98	48
ResNet50	SGDm	0.00005	84.96	42
	Adam	0.00005	92.48	45
	RMSprop	0.00005	91.73	59
	SGDm	0.0001	89.49	42
	Adam	0.0001	94.74	45
	RMSprop	0.0001	95.49	57

The Adam optimizer was observed to have provided the highest average classification accuracy of 93.12% followed by RMSProp at 89.8% and SGDm at 84.88%. Adam optimizer is a first-order gradient-based optimization method with a stochastic function considered suitable for direct application to classification models with large data and parameters [76]. Furthermore, a 0.0001 learning rate was also discovered to have generally led to higher accuracy with an average value of 90.42% while 0.00005 had 87.9%. This is consistent with the findings of Hendrawan [77] that a learning rate of 0.0001 produced better classification accuracy compared to 0.00005 for tempeh quality.

The training process for the six best models presented in Fig. 7 showed an increase in the classification accuracy as the iterations progressed. Different patterns of accuracy graphs were also obtained using the RMSprop optimizer and the validation value was observed to be increasing but fluctuating. Meanwhile, the Adam optimizer, specifically for the ResNet50 model with a learning rate of 0.0001, showed a

stable training and validation process. Adam and its modification are more preferred approaches in neural networks. Due to its exponential moving averages for both gradient and squared gradient, which significantly improve neural network training, Adam is a well-liked optimizer. These methods yield additional information about the global minimum by forcing optimization through the use of moment estimate. Furthermore, this improvement enhances the accuracy of object categorization and pattern recognition. [78]. The performance graphs for training and validation showed a rapid progression in the initial epochs, followed by convergence in the subsequent epochs, resulting in an accuracy value that approached an impressive 96.42%. The loss value consistently decreased as the iterations increased, signifying the effectiveness of the training process. The six top-performing CNN models exhibited comparable patterns, with the loss value converging towards 0. These six top performing models showed remarkable performance in classifying aflatoxin contamination levels using pre-trained networks as indicated in Fig. 8.

The determination of the best results from the training and validation process was followed by the evaluation of the CNN models' performance using the testing dataset. The confusion matrix results in Fig. 9 showed that the average accuracy of the testing dataset was 96.6% and this was considered high for the classification of the aflatoxin contamination level in cocoa beans. A confusion matrix is a matrix representation of the prediction summary. Through a comparison of the predicted and true classes, it demonstrates the number of correct and wrong predictions for each class. This information may be utilized to make decisions and optimize our

algorithms [79]. These results were comparable to the testing accuracy of 99% recorded by Albarrak [80] for date fruit classification. The CNN models were able to accurately classify 100% without errors for the "free of aflatoxin" and "contaminated above the limit" categories but had an error of 10% for the "contaminated below the limit" by misclassifying cocoa beans as contaminated above the limit. This means the models classified the cocoa beans contaminated below the limit with an accuracy of 90%. Even though there is misclassification, this type of miscalculation can still be tolerated compared to the false negative type of miscalculation, namely miscalculation of aflatoxin-contaminated cocoa beans categorized as aflatoxin-free, which will have a negative impact on ensuring food safety.

This high accuracy means the constructed CNN models can effectively classify the aflatoxin contamination level in cocoa beans as "free of aflatoxin," "contaminated below the limit," and "contaminated above the limit". Based on the results obtained above, CNN models and fluorescence images can be combined in the future to serve as an alternative method to classify the level of aflatoxin contamination in cocoa beans. This approach can also offer better accuracy and cost efficiency than existing methods. The combination of CNN models and fluorescence images is expected to minimize the impact of aflatoxin on health and losses in the cacao trade. However, this research still uses an acquisition unit on immovable objects, so further research is needed to develop an acquisition system with an acquisition unit that can record images using a motorized stage so that it approaches applications in quality control in the cacao industry.

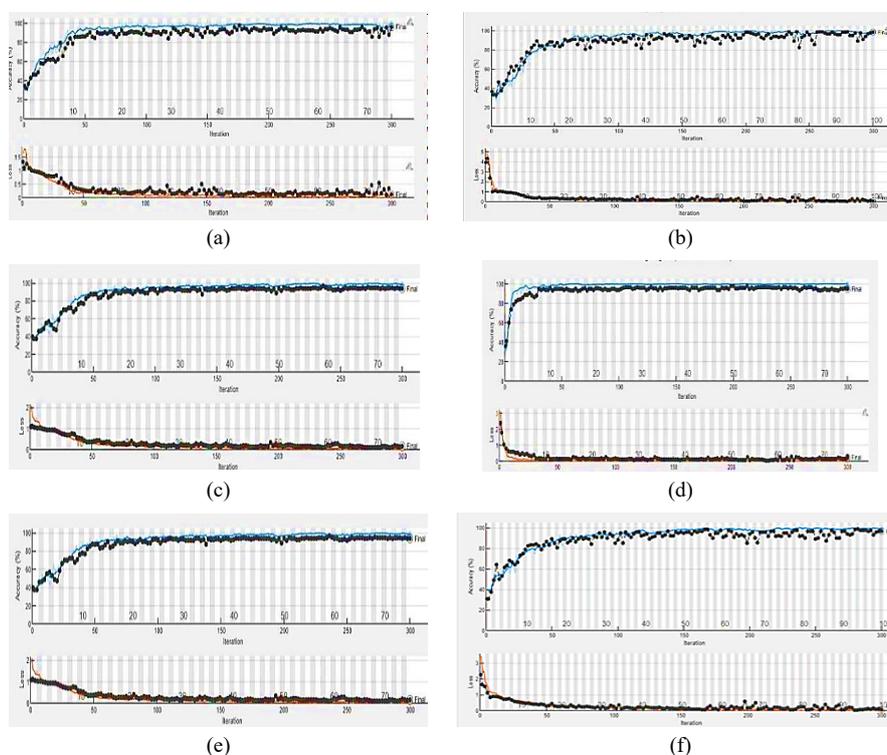


Fig. 8. Performance of the six best CNN models for aflatoxin contamination level classification using pre-trained networks: (a) GoogLeNet (optimizer = Adam, learning rate = 0.0001), (b) AlexNet (optimizer = optimizer = Adam, learning rate = 0.0001), (c) ResNet50 (optimizer = RSMprop, learning rate = 0.0001), (d) ResNet50 (optimizer = Adam, learning rate = 0.0001) (e) GoogLeNet (optimizer = Adam, learning rate = 0.00005) (f) SqueezeNet (optimizer = Adam, learning rate = 0.0001)

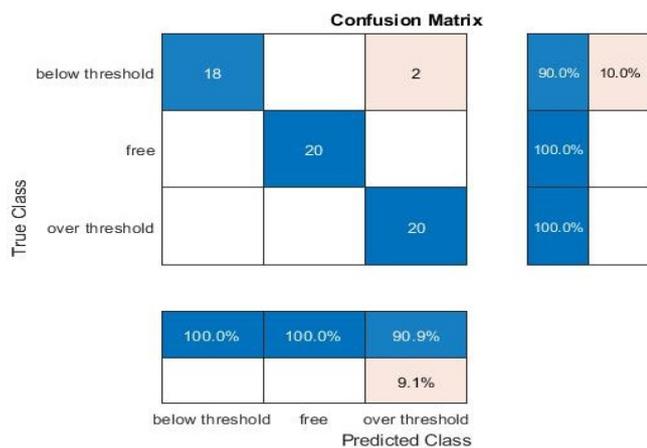


Fig. 9. Confusion matrix for the aflatoxin level classification in cocoa beans using fluorescence images

IV. CONCLUSION

In conclusion, this study demonstrated the effectiveness of utilizing pre-trained CNN models in combination with fluorescence images to accurately classify aflatoxin contamination levels in cocoa beans. Through a comparative analysis of GoogLeNet, SqueezeNet, AlexNet, and ResNet50, it was determined that the GoogLeNet model, optimized with Adam and a learning rate of 0.0001, achieved remarkable classification accuracy, with a validation accuracy of 96.42% and a testing accuracy of 96%. These results highlight the potential of deep learning techniques and image-based approaches for addressing the crucial issue of aflatoxin contamination in cocoa beans.

The success of this study opens up avenues for further research and applications. The proposed method offers a non-invasive and cost-effective means to assess the safety of cocoa beans, aiding in maintaining the quality and value of this essential commodity. Moreover, the use of fluorescence imaging holds promises for early detection of contamination, contributing to proactive interventions in the food supply chain. Future research could explore the scalability of this approach across different varieties of cocoa beans and under varying environmental conditions.

By integrating cutting-edge technology with the agriculture and food safety sectors, this study contributes to the ongoing efforts to ensure food security, trade compliance, and public health. As the field of deep learning continues to evolve, it is anticipated that innovative solutions like this will further revolutionize food quality assessment and safety assurance.

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REFERENCES

- [1] M. Eskola, G. Kos, C. T. Elliott, J. Hajšlová, S. Mayar, and R. Krška, "Worldwide contamination of food-crops with mycotoxins: Validity of the widely cited 'FAO estimate' of 25%," *Crit. Rev. Food Sci. Nutr.*, vol. 60, no. 16, pp. 2773–2789, 2020, doi: 10.1080/10408398.2019.1658570.
- [2] P. Kumar, D. K. Mahato, M. Kamle, T. K. Mohanta, and S. G. Kang, "Aflatoxins: A global concern for food safety, human health and their management," *Front. Microbiol.*, vol. 7, pp. 1–10, 2017, doi: 10.3389/fmicb.2016.02170.
- [3] J. Boonen, S. V. Malysheva, L. Tavevner, J. D. D. Mavungu, S. De Saeger, and B. D. Spiegeleer, "Human skin penetration of selected model mycotoxins," *Toxicology*, vol. 301, no. 1–3, pp. 21–32, 2012, doi: 10.1016/j.tox.2012.06.012.
- [4] S. Mohamed, Y. Sadam, R. Malaka, S. Baco, and J. Mustabi, "Aflatoxin M1 in Milk: Occurrence and Its Risk Association: A Review," *Hasanuddin Journal of Animal Science (HAJAS)*, vol. 4, no. 2, pp. 68–81, 2022, doi: 10.20956/hajas.v4i2.22097.
- [5] S. Marchese, A. Polo, A. Ariano, S. Velotto, S. Costantini, and L. Severino, "Aflatoxin B1 and M1: Biological Properties and Their Involvement in Cancer Development," *Toxins*, vol. 10, no. 6, 2018, doi: 10.3390/toxins10060214.
- [6] D. Githang'a, O. Anzala, C. Mutegi, and A. Agweyu, "The effects of exposures to mycotoxins on immunity in children: A systematic review," *Curr. Probl. Pediatr. Adolesc. Health Care*, vol. 49, no. 5, pp. 109–116, 2019, doi: 10.1016/j.cppeds.2019.04.004.
- [7] M. Mahfuz *et al.*, "Chronic Aflatoxin Exposure and Cognitive and Language Development in Young Children of Bangladesh: A Longitudinal Study," *Toxins*, vol. 14, no. 12, 2022, doi: 10.3390/toxins14120855.
- [8] R. Daou *et al.*, "Aflatoxin B1 Occurrence in Children under the Age of Five's Food Products and Aflatoxin M1 Exposure Assessment and Risk Characterization of Arab Infants through Consumption of Infant Powdered Formula: A Lebanese Experience," *Toxins*, vol. 14, no. 5, pp. 1–13, 2022, doi: 10.3390/toxins14050290.
- [9] J. Yu, "Current Understanding on Aflatoxin Biosynthesis and Future Perspective in Reducing Aflatoxin Contamination," *Toxins*, vol. 4, no. 11, pp. 1024–1057, 2012, doi: 10.3390/toxins4111024.
- [10] P. N. A. Adriansyah, W. P. Rahayu, H. D. Kusumaningrum, and O. Kawamura, "Aflatoxin M1 reduction by microorganisms isolated from kefir grains," *Int. Food Res. J.*, vol. 29, no. 1, pp. 78–85, 2022, doi: 10.47836/iftj.29.1.09.
- [11] O. Ketney, "Food Safety Legislation Regarding Of Aflatoxins Contamination," *ACTA Univ. Cibiniensis*, vol. 67, no. 1, pp. 149–154, 2015, doi: 10.1515/aucts-2015-0081.
- [12] European Commission, "COMMISSION IMPLEMENTING REGULATION (EU) No 884/2014 of 13 August 2014 imposing special conditions governing the import of certain feed and food from certain third countries due to contamination risk by aflatoxins and repealing Regulation (EC) No 1152/2009," *Off. J. Eur. Union*, vol. 242, no. 14, pp. 4–19, 2014.
- [13] P. Mounjouenpou, D. Gueule, A. Fontana-Tachon, B. Guyot, P. R. Tondje, and J. P. Guiraud, "Filamentous fungi producing ochratoxin during cocoa processing in Cameroon," *Int. J. Food Microbiol.*, vol. 121, no. 2, pp. 234–241, 2008, doi: 10.1016/j.ijfoodmicro.2007.11.017.
- [14] V. Vivek, S. Bermúdez, and C. Larrea, "Global Market Report: Cocoa," *Exch. Organ. Behav. Teach. J.*, pp. 1–12, 2019.
- [15] P. N. Pires *et al.*, "Aflatoxins and ochratoxin A: occurrence and contamination levels in cocoa beans from Brazil," *Food Addit. Contam. - Part A Chem. Anal. Control. Expo. Risk Assess.*, vol. 36, no. 5, pp. 815–824, 2019, doi: 10.1080/19440049.2019.1600749.
- [16] A. Hassoun and R. Karoui, "Quality evaluation of fish and other seafood by traditional and nondestructive instrumental methods: Advantages and limitations," *Critical Reviews in Food Science and Nutrition*, vol. 57, no. 9, pp. 1976–1998, 2017.
- [17] C. S. Alvarez *et al.*, "Aflatoxin B 1 exposure and liver cirrhosis in Guatemala: A case-control study," *BMJ Open Gastroenterol.*, vol. 7, no. 1, pp. 1–7, 2020, doi: 10.1136/bmjgast-2020-000380.
- [18] T. M. Osaili *et al.*, "Occurrence of aflatoxins in nuts and peanut butter imported to UAE," *Heliyon*, vol. 9, no. 3, p. e14530, 2023, doi: 10.1016/j.heliyon.2023.e14530.
- [19] L. Fang *et al.*, "Occurrence and exposure assessment of aflatoxins in Zhejiang province, China," *Environ. Toxicol. Pharmacol.*, vol. 92, p. 103847, 2022, doi: 10.1016/j.etap.2022.103847.
- [20] I. Polisenka, "Identification of Fusarium damaged wheat kernels using image analysis," vol. 59, no. 5, pp. 125–130, 2011, doi: 10.11118/actaun201159050125.

- [21] C. A. Mallmann, A. O. Mallmann, and D. Tyska, "Survey of Mycotoxin in Brazilian Corn by NIR Spectroscopy-Year 2019," *Glob. J. Nutri. Food Sci.*, vol. 3, pp. 1–7, 2020, doi: 10.33552/GJNFS.2020.03.000552.
- [22] X. Tian, C. Zhang, J. Li, S. Fan, Y. Yang, and W. Huang, "Detection of early decay on citrus using LW-NIR hyperspectral reflectance imaging coupled with two-band ratio and improved watershed segmentation algorithm," *Food Chem.*, vol. 360, p. 130077, 2021, doi: 10.1016/j.foodchem.2021.130077.
- [23] H. Kaya-celiker, P. K. Mallikarjunan, and A. Kaaya, "Mid-infrared spectroscopy for discrimination and classification of *Aspergillus* spp. contamination in peanuts," *Food Control*, vol. 52, pp. 103–111, 2015, doi: 10.1016/j.foodcont.2014.12.013.
- [24] M. Sieger, G. Kos, M. Sulyok, M. Godejohann, R. Krska, and B. Mizaikoff, "Portable infrared laser spectroscopy for on-site mycotoxin analysis," *Sci. Rep.*, vol. 7, 2017, doi: 10.1038/srep44028.
- [25] S. Thirupathi, D. S. Jayas, N. D. G. White, P. G. Fields, and T. Gräfenhan, "Near-Infrared (NIR) hyperspectral imaging: theory and applications to detect fungal infection and mycotoxin contamination in food products" *Indian Journal of Entomology*, vol. 78, pp. 91-99, 2016, doi: 10.5958/0974-8172.2016.00029.8.
- [26] J. Roberts, A. Power, J. Chapman, S. Chandra, and D. Cozzolino, "A short update on the advantages, applications and limitations of hyperspectral and chemical imaging in food authentication," *Appl. Sci.*, vol. 8, no. 4, 2018, doi: 10.3390/app8040505.
- [27] Z. Du, N. Ali, and B. Ashraf, "X-ray computed tomography for quality inspection of agricultural products: A review," *Food science & nutrition*, vol. 7, no. 10, pp. 3146–3160, 2019, doi: 10.1002/fsn3.1179.
- [28] J. G. Tallada, D. T. Wicklow, T. C. Pearson, and P. R. Armstrong, "Detection of Fungus-Infected Corn Kernels using Near-Infrared Reflectance Spectroscopy and Color Imaging," *Trans. ASABE*, vol. 54, pp. 1151–1158, 2011.
- [29] X. Qi, J. Jiang, X. Cui, and D. Yuan, "Identification of fungi-contaminated peanuts using hyperspectral imaging technology and joint sparse representation model," *J. Food Sci. Technol.*, vol. 56, no. 7, pp. 3195–3204, 2019, doi: 10.1007/s13197-019-03745-2.
- [30] T. Manojlović, T. Tomanić, I. Štajduhar, and M. Milanić, "Rapid extraction of skin physiological parameters from hyperspectral images using machine learning," *Appl. Intell.*, vol. 53, no. 13, pp. 16519–16539, 2023, doi: 10.1007/s10489-022-04327-0.
- [31] P. Cozzini, G. Ingleto, R. Singh, and C. Dall'Asta, "Mycotoxin detection plays 'cops and robbers': Cyclodextrin chemosensors as specialized police?," *Int. J. Mol. Sci.*, vol. 9, no. 12, pp. 2474–2494, 2008, doi: 10.3390/ijms9122474.
- [32] A. Kamilaris and F. X. Prenafeta-Boldú, "A review of the use of convolutional neural networks in agriculture," *J. Agric. Sci.*, vol. 156, no. 3, pp. 312–322, 2018, doi: 10.1017/S0021859618000436.
- [33] Z. Hruska, H. Yao, R. Kincaid, R. L. Brown, D. Bhatnagar, and T. E. Cleveland, "Temporal effects on internal fluorescence emissions associated with aflatoxin contamination from corn kernel cross-sections inoculated with toxigenic and atoxigenic *Aspergillus flavus*," *Front. Microbiol.*, vol. 8, pp. 1–10, 2017, doi: 10.3389/fmicb.2017.01718.
- [34] V. Rotich, D. F. Al Riza, F. Giametta, T. Suzuki, Y. Ogawa, and N. Kondo, "Thermal oxidation assessment of Italian extra virgin olive oil using an UltraViolet (UV) induced fluorescence imaging system," *Spectrochim. Acta - Part A Mol. Biomol. Spectrosc.*, vol. 237, p. 118373, 2020, doi: 10.1016/j.saa.2020.118373.
- [35] A. Singla, L. Yuan, and T. Ebrahimi, "Food/non-food image classification and food categorization using pre-trained GoogLeNet model," in *Proceedings of the 2nd International Workshop on Multimedia Assisted Dietary Management*, pp. 3–11, 2016, doi: 10.1145/2986035.2986039.
- [36] W. Rahman, M. G. G. Faruque, K. Rokhsana, A. H. M. S. Sadi, M. M. Rahman, and M. M. Azad, "Multiclass blood cancer classification using deep CNN with optimized features," *Array*, vol. 18, p. 100292, 2023, doi: 10.1016/j.array.2023.100292.
- [37] T. Nishi, S. Kurogi, and K. Matsuo, "Grading fruits and vegetables using RGB-D images and convolutional neural network," *2017 IEEE Symp. Ser. Comput. Intell. SSCI 2017 - Proc.*, vol. 2018 pp. 1–6, 2018, doi: 10.1109/SSCI.2017.8285278.
- [38] A. Sedik, A. Abohany, K. Sallam, K. Munasinghe, and T. Medhat, "Deep fake news detection system based on concatenated and recurrent modalities," *Expert Syst. Appl.*, vol. 208, p. 117953, 2022, doi: 10.1016/j.eswa.2022.117953.
- [39] Y. Hendrawan *et al.*, "AlexNet convolutional neural network to classify the types of Indonesian coffee beans," *IOP Conf. Ser. Earth Environ. Sci.*, vol. 905, no. 1, 2021, doi: 10.1088/1755-1315/905/1/012059.
- [40] M. Momeny, A. A. Neshat, A. Jahanbakhshi, M. Mahmoudi, Y. Ampatzidis, and P. Radeva, "Grading and fraud detection of saffron via learning-to-augment incorporated Inception-v4 CNN," *Food Control*, vol. 147, p. 109554, 2023, doi: 10.1016/j.foodcont.2022.109554.
- [41] K. Wei *et al.*, "Classification of Tea Leaves Based on Fluorescence Imaging and Convolutional Neural Networks," *Sensors (Basel)*, vol. 22, no. 20, 2022, doi: 10.3390/s22207764.
- [42] T. Tanaka *et al.*, "An application of liquid chromatography and mass spectrometry for determination of aflatoxins," *Mycotoxins*, vol. 52, no. 2, pp. 107–113, 2002, doi: 10.2520/myco.52.107.
- [43] J. Xie, N. He, L. Fang, and A. Plaza, "Scale-free convolutional neural network for remote sensing scene classification," *IEEE Trans. Geosci. Remote Sens.*, vol. 57, no. 9, pp. 6916–6928, 2019, doi: 10.1109/TGRS.2019.2909695.
- [44] V. Bhole and A. Kumar, "Analysis of Convolutional Neural Network Using Pre-Trained Squeezenet Model for Classification of Thermal Fruit Images," *ICT Compet. Strateg.*, pp. 759–768, 2020, doi: 10.1201/9781003052098-80.
- [45] N. Jamil, S. R. Roslan, R. Hamzah, and I. Ramli, "Food recognition of Malaysian meals for the management of Calorie intake," *Int. J. Adv. Trends Comput. Sci. Eng.*, vol. 8, pp. 355–360, 2019, doi: 10.30534/ijatcse/2019/5281.62019.
- [46] S. E. Abdallah, W. M. Elmessery, M. Y. Shams, N. S. A. Al-Sattary, A. A. Abohany, and M. Thabet, "Deep Learning Model Based on ResNet-50 for Beef Quality Classification," *Inf. Sci. Lett.*, vol. 12, no. 1, pp. 289–297, 2023, doi: 10.18576/isl/120124.
- [47] P. Rybacki, J. Niemann, K. Bahcevandziev, and K. Durczak, "Convolutional Neural Network Model for Variety Classification and Seed Quality Assessment of Winter Rapeseed," *Sensors*, vol. 23, no. 5, 2023, doi: 10.3390/s23052486.
- [48] S. Jiang, K. Guo, J. Liao, and G. Zheng, "Solving Fourier ptychographic imaging problems via neural network modeling and TensorFlow," *Biomed. Opt. Express*, vol. 9, no. 7, p. 3306, 2018, doi: 10.1364/boe.9.003306.
- [49] J. Jepakoch, D. M. Mugo, B. K. Kenduiyo, and E. C. Too, "The Effect of Adaptive Learning Rate on the Accuracy of Neural Networks," *Int. J. Adv. Comput. Sci. Appl.*, vol. 12, no. 8, pp. 736–751, 2021, doi: 10.14569/IJACSA.2021.0120885.
- [50] I. Kandel and M. Castelli, "The effect of batch size on the generalizability of the convolutional neural networks on a histopathology dataset," *ICT Express*, vol. 6, no. 4, pp. 312–315, 2020, doi: 10.1016/j.icte.2020.04.010.
- [51] S. Mezzah and A. Tari, "Practical hyperparameters tuning of convolutional neural networks for EEG emotional features classification," *Intell. Syst. with Appl.*, vol. 18, p. 200212, 2023, doi: 10.1016/j.iswa.2023.200212.
- [52] G. Habib and S. Qureshi, "Optimization and acceleration of convolutional neural networks: A survey," *J. King Saud Univ. - Comput. Inf. Sci.*, vol. 34, no. 7, pp. 4244–4268, 2022, doi: 10.1016/j.jksuci.2020.10.004.
- [53] S. Sarraf and G. Tofghi, "Classification of Alzheimer's Disease Structural MRI Data by Deep Learning Convolutional Neural Networks," *arXiv preprint arXiv:1607.06583*, 2016.
- [54] X. Tang, H. Zhang, N. Zhang, H. Yan, F. Tang, and W. Zhang, "Dropout Rate Prediction of Massive Open Online Courses Based on Convolutional Neural Networks and Long Short-Term Memory Network," *Mob. Inf. Syst.*, vol. 2022, p. 8255965, 2022, doi: 10.1155/2022/8255965.
- [55] A. Lopez-del Rio, M. Martin, A. Perera-Lluna, and R. Saidi, "Effect of sequence padding on the performance of deep learning models in archaeal protein functional prediction," *Sci. Rep.*, vol. 10, no. 1, pp. 1–14, 2020, doi: 10.1038/s41598-020-71450-8.
- [56] Y. Q. Yang, P. S. Wang, and Y. Liu, "Interpolation-Aware Padding for 3D Sparse Convolutional Neural Networks," *Proc. IEEE Int. Conf. Comput. Vis.*, pp. 7447–7455, 2021, doi: 10.1109/ICCV48922.2021.00737.
- [57] K. Shaheed *et al.*, "DS-CNN: A pre-trained Xception model based on

- depth-wise separable convolutional neural network for finger vein recognition," *Expert Syst. Appl.*, vol. 191, p. 116288, 2022, doi: <https://doi.org/10.1016/j.eswa.2021.116288>.
- [58] Q. Li, L. Li, W. Wang, Q. Li, and J. Zhong, "A comprehensive exploration of semantic relation extraction via pre-trained CNNs," *Knowledge-Based Syst.*, vol. 194, p. 105488, 2020, doi: <https://doi.org/10.1016/j.knsys.2020.105488>.
- [59] E. Priesterjahn, R. Geisen, and M. Schmidt-heydt, "Influence of Light and Water Activity on Growth and Mycotoxin Formation of Selected Isolates of *Aspergillus flavus* and *Aspergillus parasiticus*," *Microorganisms*, vol. 8, no. 12, pp. 1–15, 2020.
- [60] R. Casquete, M. J. Benito, M. D. G. Córdoba, and S. Ruiz-moyano, "The growth and aflatoxin production of *Aspergillus flavus* strains on a cheese model system are influenced by physicochemical factors," *J. Dairy Sci.*, vol. 100, no. 9, pp. 6987–6996, 2017, doi: [10.3168/jds.2017-12865](https://doi.org/10.3168/jds.2017-12865).
- [61] T. Furukawa and S. Sakuda, "Inhibition of aflatoxin production by paraquat and external superoxide dismutase in *Aspergillus flavus*," *Toxins*, vol. 11, no. 2, p. 107, doi: [10.3390/toxins11020107](https://doi.org/10.3390/toxins11020107).
- [62] J. Liu *et al.*, "Effects of Nutrients in Substrates of Different Grains on Aflatoxin B 1 Production by *Aspergillus flavus*," *BioMed Research International*, vol. 2016, 2016.
- [63] P. Chang, S. S. T. Hua, S. B. L. Sarreal, and R. W. Li, "Suppression of Aflatoxin Biosynthesis in *Aspergillus flavus* by 2-Phenylethanol Is Associated with Stimulated Growth and Decreased Degradation of Branched-Chain Amino Acids," *Toxins*, vol. 7, no. 10, pp. 3887–3902, 2015, doi: [10.3390/toxins7103887](https://doi.org/10.3390/toxins7103887).
- [64] E. Tumukunde *et al.*, "Osmotic-Adaptation Response of sakA/hogA Gene to Aflatoxin Biosynthesis, Morphology Development and Pathogenicity in *Aspergillus flavus*," *Toxins*, vol. 11, no. 1, pp. 1–20, 2019, doi: [10.3390/toxins11010041](https://doi.org/10.3390/toxins11010041).
- [65] Y. Wang, Y. Zhou, Y. Qin, and L. Wang, "Effect of environmental factors on the aflatoxin production by *Aspergillus flavus* during storage in upland rice seed using response surface methodology," *LWT*, vol. 169, p. 113977, 2022, doi: [10.1016/j.lwt.2022.113977](https://doi.org/10.1016/j.lwt.2022.113977).
- [66] S. Biotrop, "The effect of temperature and relative humidity for *Aspergillus flavus* BIO 2237 growth and aflatoxin production on soybeans," *International Food Research Journal*, vol. 22, no. 1, pp. 82–87, 2015.
- [67] A. Kumar, H. Pathak, and S. Bhadauria, "Aflatoxin contamination in food crops: causes, detection, and management: a review," *Food Production, Processing and Nutrition*, vol. 3, pp. 1-9, 2021.
- [68] G. Winter, C. D. Todd, M. Trovato, G. Forlani, and D. Funck, "Physiological implications of arginine metabolism in plants," *Frontiers in plant science*, vol. 6, p. 534, 2015, doi: [10.3389/fpls.2015.00534](https://doi.org/10.3389/fpls.2015.00534).
- [69] B. Wang *et al.*, "Effects of nitrogen metabolism on growth and aflatoxin biosynthesis in *Aspergillus flavus*," *J. Hazard. Mater.*, vol. 324, pp. 691–700, 2017, doi: [10.1016/j.jhazmat.2016.11.043](https://doi.org/10.1016/j.jhazmat.2016.11.043).
- [70] M. Scarpari *et al.*, "Lipids in *Aspergillus flavus* -maize interaction," *Frontiers in Microbiology*, vol. 5, pp. 1–9, 2014, doi: [10.3389/fmicb.2014.00074](https://doi.org/10.3389/fmicb.2014.00074).
- [71] I. K. Cigić and H. Prosen, "An overview of conventional and emerging analytical methods for the determination of mycotoxins," *Int. J. Mol. Sci.*, vol. 10, no. 1, pp. 62–115, 2009, doi: [10.3390/ijms10010062](https://doi.org/10.3390/ijms10010062).
- [72] J. Pawliszyn, *Comprehensive sampling and sample preparation: analytical techniques for scientists*. Academic Press, 2012.
- [73] J. Gao, L. Zhao, J. Li, L. Deng, J. Ni, and Z. Han, "Aflatoxin rapid detection based on hyperspectral with 1D-convolution neural network in the pixel level," *Food Chem.*, vol. 360, p. 129968, 2021, doi: [10.1016/j.foodchem.2021.129968](https://doi.org/10.1016/j.foodchem.2021.129968).
- [74] K. Liu, "Comparison of different Convolutional Neural Network models on Fruit 360 Dataset," *Highlights in Science, Engineering and Technology*, vol. 34, pp. 85–94, 2023.
- [75] L. Migus, J. Salomon, and P. Gallinari, "Stability of implicit neural networks for long-term forecasting in dynamical systems," *arXiv preprint arXiv:2305.17155*, 2023.
- [76] M. Reyad, A. M. Sarhan, and M. Arafa, "A modified Adam algorithm for deep neural network optimization," *Neural Computing and Applications*, pp. 1-18, 2023.
- [77] Y. Hendrawan *et al.*, "Classification of soybean tempe quality using deep learning," *IOP Conf. Ser. Earth Environ. Sci.*, vol. 924, no. 1, 2021, doi: [10.1088/1755-1315/924/1/012022](https://doi.org/10.1088/1755-1315/924/1/012022).
- [78] R. Abdulkadirov, P. Lyakhov, and N. Nagornov, "Survey of Optimization Algorithms in Modern Neural Networks," *Mathematics*, vol. 11, no. 11, pp. 1–37, 2023, doi: [10.3390/math11112466](https://doi.org/10.3390/math11112466).
- [79] D. Valero-Carreras, J. Alcaraz, and M. Landete, "Comparing two SVM models through different metrics based on the confusion matrix," *Comput. Oper. Res.*, vol. 152, p. 106131, 2023, doi: [10.1016/j.cor.2022.106131](https://doi.org/10.1016/j.cor.2022.106131).
- [80] K. Albarrak, Y. Gulzar, Y. Hamid, A. Mehmood, and A. B. Soomro, "A Deep Learning-Based Model for Date Fruit Classification," *Sustain.*, vol. 14, no. 10, 2022, doi: [10.3390/su14106339](https://doi.org/10.3390/su14106339).