Genetic Diversity and Relationship of Durian (Durio spp.) Germplasm Based on the Internal Transcribed Spacer (ITS) Region: In Silico Analysis

https://doi.org/10.18196/pt.v11i1.13649

Dindin Hidayatul Mursyidin*, Muhammad Irfan Makruf, Muhammad Fitri, Nico Aliannur

Laboratory of Genetics and Molecular Biology, Faculty of Mathematics and Natural Sciences, University of Lambung Mangkurat, Jl. A. Yani Km. 36, Banjarbaru, South Kalimantan 70714 Indonesia

*Corresponding author, email: dindinhm@gmail.com

ABSTRACT

Durian (Durio spp.) is a germplasm with a relatively high species diversity, with an estimated 27 species worldwide. However, the existence of several species has been threatened. This study aimed to reconstruct the DNA barcode of the durian and its relatives (Durio spp.) and analyze the genetic diversity and its relationship based on the internal transcribed spacer (ITS) region. Sixteen sequences of durians ITS were collected from GenBank (NCBI) and analyzed in silico using the BLAST, MultAlin, and MEGA-X software, then reconstructed phylogenetically by the UPGMA and Maximum Likelihood methods. The results show that the ITS region of Durio spp. has a base length of about 702 bp, where several mutations occur, substitution (transversion and transition) and indel (insertion and deletion). At the nucleotide level, this germplasm shows a relatively high diversity of 0.065. The cluster analyses (UPGMA and Maximum Likelihood) can separate this germplasm into four clusters and five main clades, respectively. In this study, D. zibethinus, the most popular species in the Durio genus, is closely related to D. lowianus and far from D. griffithii. This information is beneficial as reference data to support durian conservation and breeding programs, locally and globally, especially in Indonesia.

Keywords: Chloroplast DNA, Breeding, Durian, Genetic diversity, Phylogenetic

ABSTRAK

Durian (Durio spp.) merupakan salah satu plasma nutfah yang memiliki keragaman spesies relatif tinggi, diperkirakan mencapai 27 spesies di seluruh dunia, namun keberadaannya telah terancam. Penelitian ini bertujuan untuk merekonstruksi DNA barcoding durian dan kerabatnya, serta menganalisis keragaman dan kekerabatan genetiknya secara in silico berdasarkan sekuen gen internal transcribed spacer (ITS). Sebanyak 16 sekuen gen ITS Durio spp. telah dikoleksi dari GenBank (NCBI) dan dianalisis secara in silico menggunakan software BLAST, MultAlin dan MEGA-X, serta direkonstruksi secara filogenetik menggunakan metode UPGMA dan Maximum Likelihood (ML). Hasil penyejajaran memperlihatkan bahwa region tersebut memiliki panjang basa sekitar 702 bp, yang didalamnya terdapat beberapa peristiwa mutasi, baik substitusi (transversi dan transisi) dan indel (insersi dan delesi). Hasil analisis lebih lanjut menunjukkan bahwa plasma nutfah ini memiliki keragaman genetik relatif tinggi, sebesar 0,065. Sementara itu, analisis UPGMA dan ML mampu memisahkan plasma nutfah Durio spp., masing-masing kedalam empat kluster dan lima klad utama. Dalam penelitian ini, D. zibethinus yang merupakan spesies paling populer dalam genus Durio memiliki kekerabatan sangat dekat dengan D. Iowianus dan berkerabat jauh dengan D. griffithii. Informasi ini diharapkan sangat bermanfaat sebagai data acuan untuk mendukung program pelestarian dan pemuliaan durian (Durio spp.), baik secara lokal dan global, terutama di Indonesia.

Kata kunci: DNA kloroplas, Pemuliaan, Durian, Keragaman genetic, Filogenetik

INTRODUCTION

ceae family, is a higher plant with a relatively high 20 species of durian are found on several large isdiversity of species (Mursyidin et al., 2022). This lands, including Kalimantan (18 species), Sumatra germplasm is estimated to reach 27 species (Kur- (7 species), Java (1 species), Bali (1 species), Sulawesi niadinata et al., 2019). It spreads widely, especially (1 species), and Maluku (1 species) (Mursyidin in the Asian region, including Cambodia, India, & Daryono, 2016). In Kalimantan, local terms Myanmar, Sri Lanka, Thailand, Vietnam, Malaysia, have named these durians, such as lahong for D.

Durian (Durio spp.), belonging to the Malva- and Indonesia (Sundari et al., 2019). In Indonesia,



open access

dulcis, kerantungan for D. oxleyanus, and lai for D. preserving the existence of endangered species. kutejensis. Several others are known as tupaloh (D. Meanwhile, breeding/cultivation activities aim to acutifolius), apun (D. excelsus), lai kuyu (D. griffithii), explore and utilize functional genes for developing and tekawai (D. lowianus) (Uji, 2004).

cally and ecologically essential values (Aziz & Jalil, port both activities. However, this activity is per-2019). For example, nine durian species have edible formed using morphological markers, which have fruits, namely D. lowianus, D. graveolens, D. kutejen- several limitations because it is time-consuming sis, D. oxleyanus, D. testudinarium, D. grandiflorus, D. and highly influenced by environmental factors dulcis, Durio excelcus, and D. zibethinus (Aziz & Jalil, 2019). Even D. zibethinus is an agricultural commodity with prominent export prospects (Cheon et al., 2017). Indonesia, for example, one of the Among the existing molecular ones, ITS is a marker biggest durian producers in the world, was able to with advantages for characterizing germplasm, inexport this fruit to several other countries, includ- cluding durian (Santoso et al., 2017). According to ing Middle Eastern countries, with a total value of Prahl et al. (2021) and Soumya & Nair (2017), this 232,000 USD in 2020 (Rizaty, 2021). In addition, gene is located between the structural ribosomal this country produced over 1.19 million metric RNA (rRNA) of a similar precursor transcript and tons of durian in the same year (Statista Research a non-functional RNA unit with a rapid evolution-Department, 2021).

also generate wood that can be useful as interior materials. In addition, the bark of several types of durians is also used in medicine, for example, D. have been studied by various molecular markers, oxleyanus and D. griffitii as malaria drugs because they contain tannin compounds (Feng et al., 2016). However, due to various human activities, such as deforestation and excessive land conversion, especially for plantations, agriculture, settlements, and industry, several durian species have been threatened (Wilcove et al., 2013).

The International Union for Conservation of Nature or IUCN (2021) states that D. acutifolius, D. dulcis, D. grandiflorus, D. kutejensis, D. pinangianus, and *D. testudinarium* are included as vulnerable, whereas D. lanceolatus is the near-threatened. Consequently, employing conservation or preserva- Biotechnology Information (NCBI). According to tion, including cultivation and breeding efforts, is Savers et al. (2019), GenBank has a comprehensive indispensable. According to Wintle et al. (2019), database of freely accessible nucleotide sequences conservation is an activity directed at saving and or formal gene descriptions. Hence, such a study

new superior cultivars (van Huylenbroeck, 2018). In general, most durian species have economi- In this case, characterization is also urgent to sup-(Mursyidin & Khairullah, 2020).

The molecular markers provide speed and high accuracy in germplasm characterization activities. ary rate. As a result, it can be used to determine Apart from producing fruit, 14 species of durian germplasm relationships at the genus, species, and subspecies levels (Qin et al., 2017).

> Previously, the genetics of durian germplasm such as RAPD (Mursyidin & Daryono, 2016; Prihatini et al., 2016; Hariyati et al., 2013), SSR, and ISSR (Ho et al., 2020; Santoso et al., 2016). However, these markers are highly subjective. In addition, poor consistency, limited repeatability, and complicated operation limit their effectiveness (Wu et al., 2021). This study aimed to reconstruct a DNA barcoding motif, as well as determine and analyze the genetic diversity and relationship of 16 durian species (Durio spp.) in silico by utilizing internal transcribed spacer (ITS) gene sequence data provided in GenBank or the National Center for

MATERIALS AND METHODS

This study was conducted from March-May 2020 *in silico* at the Laboratory of Genetics and Molecular Biology, Faculty of Mathematics and Natural Sciences, University of Lambung Mangkurat. This research used internal transcribed spacer (ITS) gene sequence data from 16 durian species (*Durio* spp.) found in GenBank or NCBI (Table 1). In general, this study includes three main activities: nucleotide sequence search and homology (similarity) analysis, nucleotide (multiple) sequence alignment, and analysis of genetic diversity and relationship of those durians obtained.

Nucleotide Sequence Search and Homology Analysis

The internal transcribed spacer (ITS) gene sequence of *D. zibethinus*, *which is* available on the GenBank or NCBI website (https://www. ncbi.nlm.nih.gov/) with accession number MF629779.1, was used as a reference in this study (Table 1). The homology (similarity) analysis with other durian species, also available on the GenBank website, was then carried out using the BLAST (Basic Local Alignment Search Tool) software. All of the durians (*Durio* spp.) ITS sequences were then copied into text (notepad) format for further analysis.

Multiple Sequence Alignments

Nucleotide (multiple) sequence alignment of the ITS durian region and its relatives (*Durio* spp.) was carried out online using the MultAlin software (<u>Babar et al., 2014</u>). The software is available online at http://multalin.toulouse.inra.fr/multalin/.

Analysis of Genetic Diversity and Relationship

Analysis of genetic diversity and the relationship of Durio spp. was performed using Molecular Evolutionary Genetics Analysis or MEGA-X software (<u>Kumar et al., 2018</u>). This analysis was started by inputting all ITS durian sequence data (text format) into the MEGA-X software. Before analysis, the sequences were first converted to fasta (.fas) or mega (.meg) format and aligned in the software. After that, genetic diversity analysis was carried out using the nucleotide diversity index (π) method (Nei & Li, 1979). Meanwhile, phylogenetic reconstruction was employed using the UPGMA and Maximum Likelihood methods (Swenson, 2019). The statistical (bootstrap) analysis was then applied to evaluate the internal branches of the phylograms (Kumar et al., 2018).

RESULTS AND DISCUSSION

Multiple Sequence Alignments

Multiple sequence alignment is one of the biological studies which is most frequently used data analysis models (Shi et al., 2021). This modeling is applied to look at the phylogeny of a whole genome, proteins, identification of horizontally transferred genes, and detection of combined sequences (Zielezinski et al., 2017). As data sequencing technology advances, the use of this modeling is increasing (Katoh et al., 2018). The increasing use of multiple sequence alignment modeling has made this area an active research topic, so more than 100 methods have been used. In multiple alignment analysis, all data will be entered from a point in a set of sequences into equivalent classes based on their respective similarities for all members of a common ancestor (Maiolo et al., 2018).

The durian and its relatives (*Durio* spp.) have a total length of ITS gene sequences of around 702 base pairs (Figure 1), in which several mutation events, both substitutions (transitions and transversions) and insertion-deletion (indel), are found. Table 2 provides detailed information about mutation events in the ITS durian sequences. There were 217 loci experiencing mutations in the durian ITS gene sequence (Figure 1 and Table 2). tion (95 loci) compared to others. Compared to is universal for different taxa. Moreover, it can be other studies, the number of mutations in durian is utilized in phylogenetic studies, molecular ecolhigher than in other species. Soumva & Nair (2017) ogy, detection, and identification of individual reported that in the ITS region of Averrhoa (L.), pathogens and non-pathogens (Zhang et al., 2021). there were only 87 loci mutations out of a total of According to Skuza et al. (2019), the ITS region has 615 bases it had. Similar other cases were shown in proven to be one of the most informative regions Anoectochilus (Thinh et al., 2020), Aquilaria (Lee et for forming genetic relationships between species al., 2017), Litsea (Fijridivanto & Murakami, 2019), in the genus. Uncaria (Zhu et al., 2020), and Zanthoxylum (Zhao et al., 2018).

Transversion was the most common type of muta- ribosomal RNA with a high evolutionary rate but

In this case, however, the deletion was the lowest found in the ITS sequence of durians (13 loci). These study results align with <u>Soumya & Nair</u> A similar result was found in fungi (<u>Zhang et al.</u>, (2017), stating that the ITS region is a part of 2021). According to Houde et al. (2019), such a

Table 1. Species of Durians used in this study, the nucleotide length, and GenBank accession number

Species	Nucleotide length (bp)	GenBank Accession Number
D. acutifolius	684	AF287700.1
D. affinis	692	AF287705.1
D. beccarianus	695	AF287707.1
D. carinatus	689	AF287708.1
D. dulcis	689	AF287713.1
D. grandiflorus	683	AF233320.1
D. graveolens	729	MF629770.1
D. griffithii	684	AF233310.1
D. kutejensis	730	MF629750.1
D. lanceolatus	686	AF287709.1
D. lowianus	688	AF287711.1
D. oblongus	692	AF287703.1
D. oxleyanus	688	AF233306.1
D. singaporensis	696	AF287701.1
D. testudinarium	694	AF287704.1
D. zibethinus*	747	MF629779.1

*Reference species

Table 2. Mutations on the ITS region of Durio spp. germplasm

Mutation type	Number of Mutation
Deletion	13
Insertion	15
Substitution-transition	94
Substitution -transversion	95
Total	217

Table 3. Information of internal transcribed spacer (ITS) of Durio spp*

Parameter	Value
Nucleotide length (bp)	684-747
Number of the polymorphic site	217
Bayesian information criteria (BIC)	4538.350
Akaike information criteria (AIC)	4320.399
Maximum likelihood value (ÌnL)	-2130.112
Transition/tranversion bias value (R)	1.010
GC content (%)	68.710
Nucleotide diversity (π)	0.065

* following Kimura 2-parameter model

Durio zib dhinus Durio lowianus Durio dukas Durio dukas Durio grawolans Durio katgensis Durio katgensis Durio katgensis Durio testudinarium Durio becarianus Durio sing aporensis Durio sing aporensis Durio graffihi Dunio grandiflorus Durio grandiflorus Durio acuifolius Durio acuifolius

Durio zib ahinus Durio okuanus Durio dulcis Durio grawolans Durio grawolans Durio carinatus Durio kutgasis Durio kutgasis Durio katanus Durio becarianus Durio offinis Durio affinis Durio grandifichis Durio grandifichis Durio grandifichis Durio acuifolius Consersus

Durio zibethinus Durio lowianus Durio dukas Durio graveolens Durio carinatus Durio katejensis Durio katejensis Durio katejensis Durio testudinarium Durio becarianus Durio sing aporensis Durio griftihi Durio grandiflorus Durio grandiflorus Durio grandiflorus Durio acuifolius Durio acuifolius

Durio zib ethinus Durio lowianus Durio dulcis Durio graveolens Durio carinatus Durio katejensis Durio katejensis Durio katejensis Durio becarianus Durio becarianus Durio sing aporensis Durio grafitiki Durio grandiflorus Durio grandiflorus Durio garantiflotus Durio garantiflotus Durio garantiflotus

Durio zibethinus Durio olovianus Durio dulcis Durio graveolens Durio graveolens Durio lanceolatus Durio lanceolatus Durio bettefensis Durio testudinarium Durio becarianus Durio singaporensis Durio singaporensis Durio grifithi Durio grandiflorus Durio grandiflorus Durio acuifolius Durio acuifolius

Durio zib ethinus Durio lowianus Durio dulcis Durio grawolens Durio carinatus Durio lanceolatus Durio kuteersis Durio kuteersis Durio testudinarium Durio becarianus Durio schongus Durio sing aporensis Durio grafithi Durio grandiflorus Durio grandiflorus Durio grandiflorus Durio acutifolius Durio acutifolius

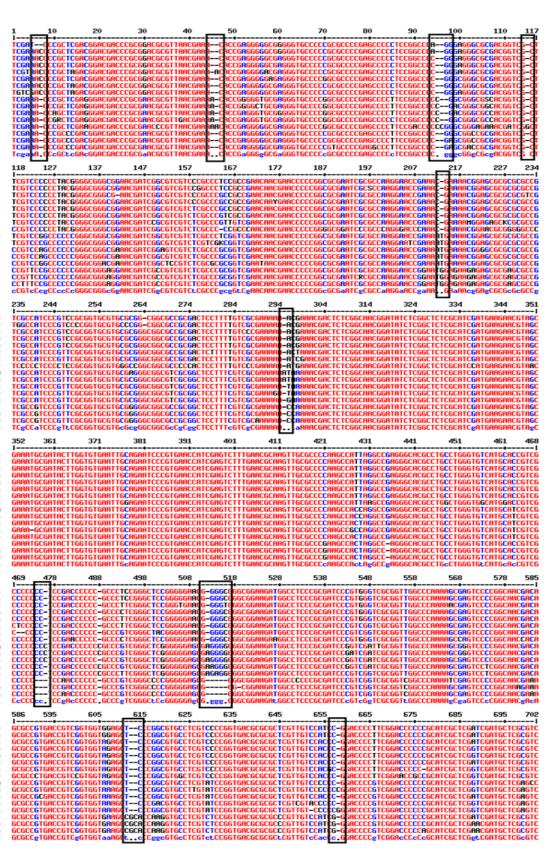


Figure 1. Multiple sequence alignment of the ITS region of *Durio* spp., showing a unique DNA barcoding motif, where mutation, like indels, present therein (close rectangle)

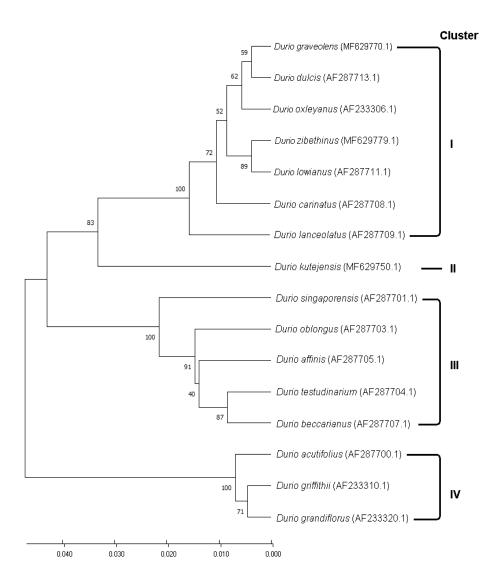


Figure 2. The genetic relationship of *Durio* spp. based on UPGMA analysis. The value on the internal branch shows the results of the bootstrap analysis 1000 replicates

mutation has a significant value in phylogenetic relations or increases the resolution of evolutionary genetic relationships between the studied taxa. Among candidate DNA barcoding regions, ITS is a non-coding region that generally shows high genetic diversity, including indel polymorphism, so it has the potential ability to be applied in species identification (Qin et al., 2017).

Genetic Diversity and Relationship of Durio spp.

Durian and its relatives (*Durio* spp.) showed relatively high genetic diversity at the nucleotide level,

recorded at 0.065 (Table 3). This genetic diversity is closely related to mutation events in the ITS durian sequences studied. Based on Table 3, the *Durio* spp. has ITS sequence character with relatively high polymorphic sites (217 loci), relatively high GC content (68.71%), and the substitution bias value (transition/transversion) is also high (1.01).

According to <u>van Dorp et al. (2020)</u>, mutations are the main factor in the emergence of genetic diversity. In other words, this phenomenon is the primary source that plays a vital role in knowing the differences and evolutionary events between

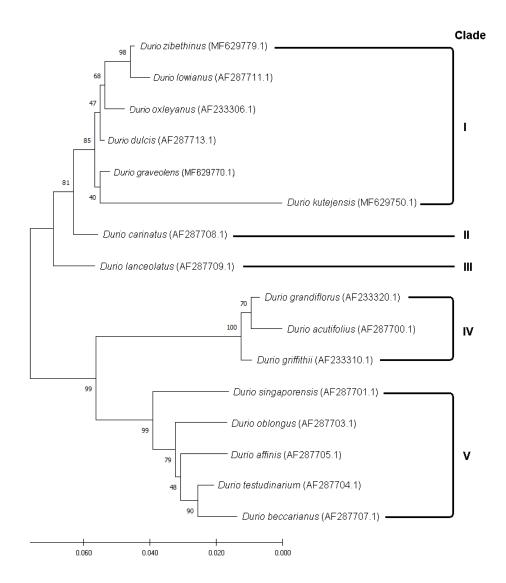


Figure 3. The genetic relationship of Durio spp. based on Maximum Likelihood (ML) analysis. The value on the internal branch shows the results of the bootstrap analysis 1000 replicates

species. In short, a mutation is an event of perma- cause variations between species. nent changes in genetic material, genes, genomes, and chromosomes (Maluszynski et al., 2017). At and Maximum Likelihood or ML (Figure 3), the the nucleotide (gene or genome) level, mutations durian germplasm (Durio spp.) was separated into include four types, namely deletions, insertions, four clusters and five main clades, respectively. transitions, and transversions. Deletion and inser- The separation is generally relatively consistent tion mutations can cause a shift in the coding of for all species that group together, either into the the nucleotide triplet. Meanwhile, transition and same cluster or clade. It is strongly related to the transversion mutations can change the amino acid bootstrap analysis results and is relatively high in composition formed (Guo et al., 2017). According the resulting internal branches of the dendrogram to Qin et al. (2017), internal transcribed spacer (Figure 2) and phylogram (Figure 3). (ITS) sequences generally have mutations that In this study, *D. zibethinus* closely related to *D.*

Based on the UPGMA method (Figure 2)

lowianus but very far from *D. griffithi* (see Figures (transversion and transition) and indel (insertion 2 and 3). This study's results align with Nyffeler and deletion). The results also show that this <u>& Baum (2000)</u> using the ITS marker, stating germplasm has a relatively high genetic diversity, that D. zibethinus is closely related to D. lowianus. amounting to 0.065. Meanwhile, the cluster analy-According to Nyffeler & Baum (2000), these two sis (UPGMA) and Maximum Likelihood (ML) can durian species have morphological similarities, separate Durio spp. into four clusters and five main such as calyx, filament arrangement, and anther clades, respectively. In this study, D. zibethinus, the architecture. Based on other studies, mainly us- most popular species in the genus Durio, is closely ing markers *ndh*F and ITS, a close relationship related to *D. lowianus* and distantly related to *D.* between D. zibethinus and D. oxleyanus was reported griffithii. This information is beneficial as reference (Nyffeler & Baum, 2001). Meanwhile, based on the data to support durian (Durio spp.) conservation ITS nr-DNA marker, the relationship between *D*. and breeding programs, both locally and globally, zibethinus and D. kutejensis was reported (Santoso especially in Indonesia. et al., 2017). Furthermore, following the UPGMA (Figure 2) and ML (Figure 3), D. carinatus and D. lanceolatus are nearly related. Naufal (2021) reported that these two durians had the same primitive character in the form of a straight pistil stalk and flowering present on the branches.

However, the relationship between these organisms can show the results of molecular evolution during a time course in the presence of genetic differences (Guerrero et al., 2019). In other words, phylogenetic trees obtained from polymorphisms in a genome, such as chloroplasts, can be used to determine the bar code of an organism and identify differences in taxonomic status and evolution, including population genetics (Nguyen et al., 2017; Xu et al., 2020). Furthermore, information on these relationships benefits the conservation of endangered species and increases plant breeding in general (<u>Bi et al., 2018</u>).

CONCLUSION

Durian germplasm (Durio spp.) shows a unique DNA barcode motif based on the internal transcribed spacer (ITS) region sequence. The alignment of these sequences shows that the ITS Durio spp. has a base length of about 702 bp, in which there are several mutation events, both substitution

ACKNOWLEDGMENTS

This study was funded partly by the Director General of Higher Education, Ministry of Education and Culture, Indonesia, through a national student competitive research grant for 2020.

REFERENCES

- Aziz, N. A. A., & Jalil, A. M. M. (2019). Bioactive compounds, nutritional value, and potential health benefits of indigenous durian (Durio zibethinus Murr.): A review. Foods, 8(3), 1-18. https:// doi.org/10.3390/foods8030096.
- Babar, M. E., Pervez, M. T., Nadeem, A., Hussain, T., & Aslam, N. (2014). Multiple sequence alignment tools: assessing performance of the underlying algorithms. Journal of Applied Environmental and Biological Sciences, 4(8S), 76-80.
- Bi, Y., Zhang, M. F., Xue, J., Dong, R., Du, Y. P., & Zhang, H. X. (2018). Chloroplast genomic resources for phylogeny and DNA barcoding: A case study on Fritillaria, Scientific Reports, 8(1), 1-12, https://doi.org/10.1038/s41598-018-19591-9.
- Cheon, S. H., Jo, S., Kim, H. W., Kim, Y. K., Sohn, J. Y. & Kim, K. J. (2017). The complete plastome sequence of Durian, Durio zibethinus L. (Malvaceae). Mitochondrial DNA Part B: Resources, 2(2), 763-764. https://doi.org/10.1080/23802359.2017.1398615.
- Feng, J., Wang, Y., Yi, X., Yang, W. & He, X. (2016). Phenolics from durian exert pronounced NO inhibitory and antioxidant activities. Journal of Agricultural and Food Chemistry, 64(21), 4273-4279. https://doi.org/10.1021/acs.jafc.6b01580
- Fijridiyanto, I. A., & Murakami, N. (2019). Evaluating the utility of external transcribed spacer (ETS) and internal transcribed spacer sequences (ITS) for phylogenetic analysis of Litsea Lam. (Lauraceae) and related genera. Buletin Kebun Raya, 22(1), 47-68.
- Guerrero, P. C., Majure, L. C., Cornejo-Romero, A., & Hernández-

Hernández, T. (2019). Phylogenetic relationships and evolutionary trends in the cactus family. *Journal of Heredity*, *110*(1), 4–21. <u>https://doi.org/10.1093/jhered/esy064</u>.

- Guo, C., McDowell, I. C., Nodzenski, M., Scholtens, D. M., Allen, A. S., Lowe, W. L., & Reddy, T. E. (2017). Transversions have larger regulatory effects than transitions. *BMC Genomics*, 18(394), 1–9. <u>https://doi.org/10.1186/s12864-017-3785-4</u>
- Hariyati, T., Kusnadi, J., & Arumingtyas, E. L. (2013). Genetic diversity of hybrid durian resulted from cross-breeding between *Durio kutejensis* and *Durio zibethinus* based on random amplified polymorphic DNAs (RAPDs). *American Journal of Molecular Biology*, 03, 153–157. <u>https://doi.org/10.4236/ajmb.2013.33020</u>.
- Ho, V. T., Ho, M. D., & Tran, T. L. (2020). Characterizing genetic variation of two popular durians (*Durio zibethinus* L.) varieties in southern Vietnam by using ISSR markers. *Bioscience Research*, 17, 3040–3049.
- Houde, P., Braun, E. L., Narula, N., Minjares, U., & Mirarab, S. (2019). Phylogenetic signal of indels and the Neoavian radiation. *Diversity*, 11(7), 1–23. <u>https://doi.org/10.3390/d11070108</u>.
- IUCN. (2021). The IUCN Red List of Threatened Species: Durio. Retrieved March 27, 2021, from <u>https://www.iucnredlist.org/</u>
- Katoh, K., Rozewicki, J. & Yamada, K.D. (2018). MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefings in Bioinformatics*, 20(4), 1160–1166. <u>https://doi.org/10.1093/bib/bbx108</u>.
- Kumar, S., Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution*, 35(6), 1547–1549. https://doi.org/10.1093/molbev/msy096.
- Kurniadinata, O. F., Wenpei, S., Zaini, A., & Rusdiansyah. (2019). Six potential superior durian plants resulted by cross breeding of D. zibethinus and D. kutejensis from East Kalimantan, Indonesia: initial identification. Journal of Tropical Horticulture, 2(2), 45. http://dx.doi.org/10.33089/jthort.v2i2.24.
- Lee, S. Y., Mohamed, R., Faridah-Hanum, I., & Lamasudin, D. U. (2018). Utilization of the internal transcribed spacer (ITS) DNA sequence to trace the geographical sources of Aquilaria malaccensis Lam. populations. Plant Genetic Resources: Characterisation and Utilisation, 16(2), 103–111. <u>https://doi.org/10.1017/</u> <u>S1479262117000016</u>
- Maiolo, M., Zhang, X., Gil, M., & Anisimova, M. (2018). Progressive multiple sequence alignment with indel evolution. BMC Bioinformatics, 19(1), 1–8. <u>https://doi.org/10.1186/s12859-018-2357-1</u>.
- Maluszynski, M., Szarejko, I., Maluszynska, J., & Szurman-Zubrzycka, M. (2017). Mutation techniques. In *Encyclopedia of Applied Plant Sciences* (Vol. 2, pp. 215–228). Elsevier Inc. <u>https://dx.doi.org/10.1016/B978-0-12-394802-6.00121-0.</u>
- Mursyidin, D. H. & Daryono, B. S. (2016). Genetic diversity of local durian (*Durio zibethinus* Murr.) cultivars of South Kalimantan's province based on RAPD markers. *AIP Conference Proceedings*, 1755(040008), 1-7. <u>https://doi.org/10.1063/1.4958483</u>.
- Mursyidin, D. H., & Khairullah, I. (2020). Genetic evaluation of tidal swamp rice from South Kalimantan, Indonesia based on the agro-morphological markers. *Biodiversitas Journal of Biological Diversity*, 21(10), 4795–4803. <u>https://doi.org/10.13057/ biodiv/d211045.</u>

- Mursyidin, D. H., Makruf, M. I., Badruzsaufari, & Noor, A. (2022). Molecular diversity of exotic durian (*Durio* spp.) germplasm: a case study of Kalimantan, Indonesia. *Journal of Genetic Engineering* and Biotechnology, 20(39), 1–13. <u>https://doi.org/10.1186/</u> s43141-022-00321-8
- Naufal, D. I. (2021). Studi filogenetika Durio di Kalimantan berdasarkan karakter morfologi bunga (Unpublished Thesis). Universitas Islam Negeri Syarif Hidayatullah.
- Nei, M. & Li, W.H. (1979). Mathematical model for studying genetic variation in terms of restriction endonucleases. *Proceedings* of the National Academy of Sciences, 76(10), 5269–5273. https://doi.org/10.1073/pnas.76.10.5269.
- Nguyen, V. B., Park, H. S., Lee, S. C., Lee, J., Park, J. Y., & Yang, T. J. (2017). Authentication markers for five major *Panax* species developed via comparative analysis of complete chloroplast genome sequences. *Journal of Agricultural and Food Chemistry*, 65(30), 6298-6306. https://doi.org/10.1021/acs.jafc.7b00925.
- Nyffeler, R. & Baum, D.A. (2000). Phylogenetic relationships of the durians (Bombacaceae-Durioneae or /Malvaceae/Helicteroideae/Durioneae) based on chloroplast and nuclear ribosomal DNA sequences. *Plant Systematics and Evolution*, 224(1–2), 55–82. https://doi.org/10.1007/BF00985266.
- Nyffeler, R. & Baum, D. A. (2001). Systematics and character evolution in *Durio* s. lat. (malvaceae/helicteroideae/durioneae or bombacaceae-durioneae). *Organisms Diversity and Evolution*, 1(3), 165–178. https://doi.org/<u>10.1078/1439-6092-00015</u>.
- Parikesit, A. A., Anurogo, D. & Putranto, R. A. (2017). Pemanfaatan bioinformatika dalam bidang pertanian dan kesehatan. *Menara Perkebunan*, 85(2), 105–115. <u>http://dx.doi.org/10.22302/iribb.</u> jur.mp.v85i2.237.
- Prahl, R. E., Khan, S., & Deo, R. C. (2021). The role of internal transcribed spacer 2 secondary structures in classifying mycoparasitic Ampelomyces. *PLoS ONE*, 16, 1-28. https://doi. org/10.1371/journal.pone.0253772.
- Prihatini, R., Ihsan, F., & Indriyani, N. L. P. (2016). Genomic profiling of F1 hybrids of durian (*Durio zibethinus*) revealed by RAPD-PCR. *Journal of Horticulture Research*, 24, 69–76. <u>https://doi. org/10.1515/johr-2016-0022.</u>
- Qin, Y., Li, M., Cao, Y., Gao, Y., & Zhang, W. (2017). Molecular thresholds of ITS2 and their implications for molecular evolution and species identification in seed plants. *Scientific Reports*, 7(1), 1–8. <u>https://doi.org/10.1038/s41598-017-17695-2</u>.
- Rizaty, M.A. (2021). National production of durian [Produksi durian nasional]. Retrieved September 27, 2021, from https://databoks.katadata.co.id/datapublish/2021/06/23/produksi-durian-di-indonesia-menurun-pada-2020.
- Santoso, P. J., Granitia, A., Indriyani, N. L. P., & Pancoro, A. (2016). Loci analysis and diversity of durian (*Durio* sp.) germplasm based on microsatellite markers. *Jurnal Hortikultur*, 26, 9–20.
- Santoso, P. J., Indriyani, N. L. P., Istianto, M., Pancoro, A., & Aryantha, I. N. P. (2017). Phylogeny of Indonesian durian (*Durio* sp.) germplasm based on polymorphism of ITS-nrDNA sequences. *Acta Horticulturae*, 1186, 35–41.
- Sayers, E. W., Mark, C., Karen, C., James, O., Kim, D. P., & Ilene, K. M. (2019). GenBank. *Nucleic Acids Research*, 47, D94–D99.
- Shi, H., Shi, H., & Xu, S. (2021). Efficient multiple sequences alignment algorithm generation via components assembly under

PAR framework. *Frontiers in Genetics*, *11*, 1–7. https://doi. org/<u>10.3389/fgene.2020.628175</u>.

- Skuza, L., Szućko, I., Filip, E., & Strzała, T. (2019). Genetic diversity and relationship between cultivated, weedy and wild rye species as revealed by chloroplast and mitochondrial DNA noncoding regions analysis. *PLoS ONE*, *14*(2), 1–21. <u>https://doi.org/10.1371/journal.pone.0213023</u>.
- Soumya, S. L. & Nair, B. R. (2017). Internal transcribed spacer (ITS) sequence analysis of nuclear ribosomal DNA (nrDNA) in Averrhoa L. International Journal of Current Research, 9(1), 45353–45359.
- Statista Research Department (2021). Production of durian in Indonesia 2011-2020. Retrieved November 05, 2021, from https://www.statista.com/statistics/706504/production-ofdurian-in-indonesia/
- Sundari, Mas'ud, A., Arumingtyas, E. L., Hakim, L., Azrianingsih, R., & Wahyudi, D. (2019). Taxonomic status of local durian (*Durio* spp.) from Ternate Island Noth Maluku based on morphological character and geographical factor. *International Journal of Conservation Science*, 10 (4), 711–720.
- Swenson, N. G. (2019). *Phylogenetic ecology: A history, critics & remodelling.* The University of Chicago Press.
- Thinh, B. B., Chac, L. D., & Thu, L. T. M. (2020). Application of internal transcribed spacer (ITS) sequences for identifying Anoectochilus setaceus Blume in Thanh Hoa, Vietnam. Proceedings on Applied Botany, Genetics and Breeding, 181(2), 108-116. https://doi.org/10.30901/2227-8834-2020-2-108-116
- Uji, T. (2004). Keanekaragaman jenis, plasma nutfah, dan potensi buah-buahan asli Kalimantan. *BioSmart*, *6*(2), 117-125.
- van Dorp, L., Acman, M., Richard, D., Shaw, L. P., Ford, C. E., Ormond, L., Owen, C. J., Pang, J., Tan, C. C. S., Boshier, F. A. T., Ortiz, A. T., & Balloux, F. (2020). Emergence of genomic diversity and recurrent mutations in SARS-CoV-2. *Infection, Genetics, and Evolution*, 83, 1–9. <u>https://doi.org/10.1016/j.meegid.2020.104351</u>.
- van Huylenbroeck J. (2018). Handbook of Plant Breeding: Ornamental crops. Springer International Publishing AG.
- Wilcove, D. S., Giam, X., Edwards, D. P., Fisher, B., & Koh, L. P. (2013). Navjot's nightmare revisited: logging, agriculture, and biodiversity in Southeast Asia. *Trends in Ecology and Evolution*, 28(9), 531–540. <u>https://doi.org/10.1016/j.tree.2013.04.005</u>.
- Wintle, B. A., Kujala, H., Whitehead, A., Cameron, A., Veloz, S., Kukkala, A., Moilanen, A., Gordon, A., Lentini, P. E., Cadenhead, N. C. R., & Bekessy, S. A. (2019). Global synthesis of conservation studies reveals the importance of small habitat patches for biodiversity. *PNAS*, *116*(3), 909–914. <u>https://doi.org/10.1073/ pnas.1813051115</u>.
- Wu, F., Ma, S., Zhou, J., Han, C., Hu, R., Yang, X., Nie, G., & Zhang, X. (2021). Genetic diversity and population structure analysis in a large collection of white clover (*Trifolium repens* L.) germplasm worldwide. *PeerJ*, 9, 1–17. <u>https://doi.org/10.7717/ peerj.11325</u>.
- Xu, J., Shen, X., Liao, B., Xu, J., & Hou, D. (2020). Comparing and phylogenetic analysis chloroplast genome of three Achyranthes species. Scientific Reports, 10(1), 1–13. <u>https://doi.org/10.1038/s41598-020-67679-y</u>.

Zhang, Y. Z., Han, Q. D., Fu, L. W., Wang, Y. X., Sui, Z. H., & Liu, Y.

G. (2021). Molecular identification and phylogenetic analysis of fungal pathogens isolated from diseased fish in Xinjiang, China. *Journal of Fish Biology*, *99*(6), 1887–1898. https://doi. org/<u>10.1111/jfb.14893.</u>

- Zhao, L. L., Feng, S. J., Tian, J. Y., Wei, A. Z., & Yang, T. X. (2018). Internal transcribed spacer 2 (ITS2) barcodes: A useful tool for identifying Chinese *Zanthoxylum*. *Applications in Plant Sciences*, 6(6), e1157. <u>https://doi.org/10.1002/aps3.1157</u>
- Zhu, S., Li, Q., Chen, S., Wang, Y., Zhou, L., Zeng, C., & Dong, J. (2018). Phylogenetic analysis of Uncaria species based on internal transcribed spacer (ITS) region and ITS2 secondary structure. *Pharmaceutical Biology*, *56*(1), 548–558. <u>https://doi.org/10.1 080/13880209.2018.1499780</u>
- Zielezinski, A., Vinga, S., Almeida, J., & Karlowski, W. M. (2017). Alignment-free sequence comparison: benefits, applications, and tools. *Genome Biology*, 18(1), 1–17. https://doi.org/10.1186/ s13059-017-1319-7.